



ACCU-SCOPE®

MANUAL

EXI-600
INVERTED RESEARCH
MICROSCOPE SERIES



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1. ATTENTION

This manual is written for the EXI-600 research inverted biological microscope. To ensure the safety, obtain optimum performance and to familiarize yourself fully with the microscope, it is strongly recommended that you read this manual carefully before operating the microscope and put this manual in a place easy to access for reference.

ACCU-SCOPE, Inc. provides safe and reliable instruments, but improper operation and the avoidance of precautions may lead to personal injury or loss of property. To ensure proper operation and optimization of the product features, please read this manual carefully before using the product. It is highly recommended to put store this manual in an easily accessible location for quick reference.

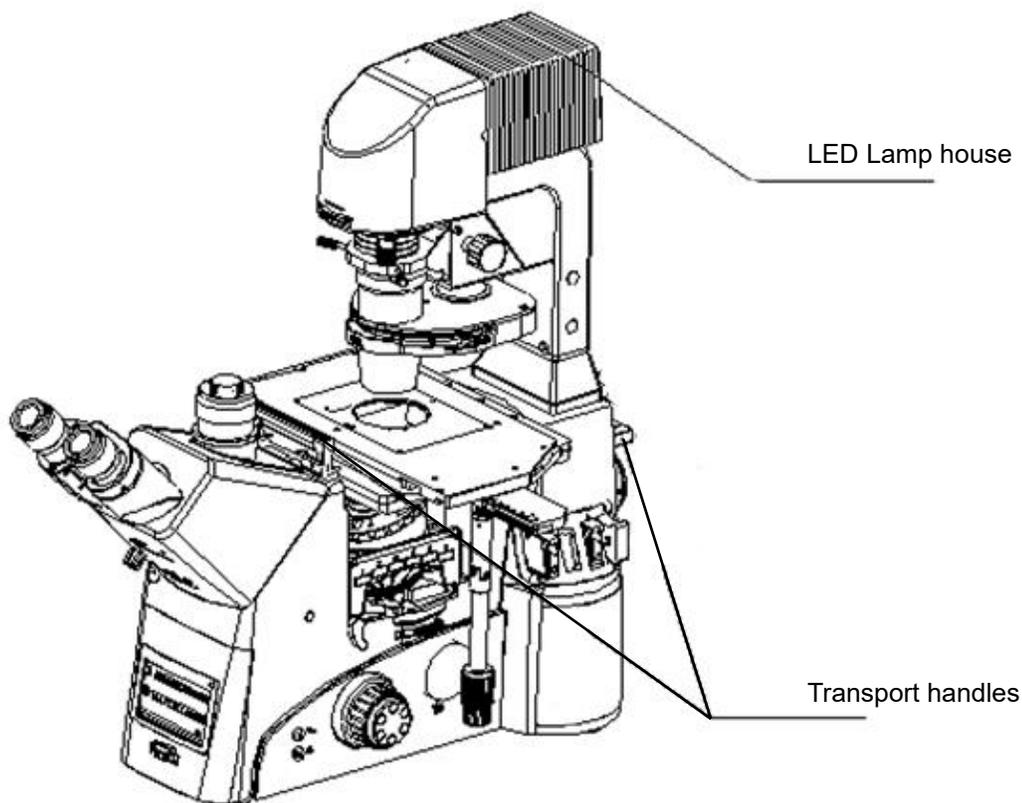
Safety notices in this manual will be identified and emphasized by the below symbols. Please pay particular attention to the sections where these symbols appear.

Symbol	Meaning
 WARNING	Symbol indicates that ignoring this alert may result in personal injury or even death.
 NOTICE	Symbol indicates that ignoring this alert may result in personal injury or loss of property.

Meaning of symbols found on the instrument.

Symbol	Meaning
	<i>Ground fault terminal</i>
	CAUTION <i>Hot surface!</i>
	CAUTION <i>Indicates hazard to the user of the instrument.</i>
	CAUTION <i>Disconnect power to the instrument before opening it.</i>

2. SAFETY PRECAUTIONS



1. Avoid placing the microscope in a location exposed to direct sunlight, with high temperature, wide temperature fluctuations, high humidity or vibration. Ensure the bench/platform is level, flat and can support the entire weight of the microscopes and any peripherals and accessories (mainframe weighs ~30kg).
2. When moving the microscope, hold the transport handles tightly and raise the microscope above the bench before moving it. Do not drag/push the microscope across the bench.
3. If liquid (e.g., culture medium, water, etc.) drips or is spilled onto the stage, objective or observation tube, immediately turn off the power supply and wipe off the solution to prevent further damage.
4. When operating, the lamp house can be considerably hot, make sure there is enough space around the lamp house for heat dissipation to avoid the heat accumulates and damage the instrument. Allow the lamp house to cool completely before covering the microscope with the dust cover.





5. Before turning on the power to the instrument, ensure the power supply is properly connected.

Before replacing the bulb or fuse, turn off the power and wait until the lamp house has cooled completely.

- ★ The fuse should have the same specifications as the original. Do not use a temporary fuse in place of a damaged/burned fuse.

6. Connect the power cables correctly to ensure the instrument is grounded in case of lightning strike.

7. Use only the included, dedicated cables.

8. Install the instrument in a location away from any acidic gasses, alkaline agents, organic solvents and any harmful substances that may damage the instrument or harm the operator.

- ★ A grounding 3-pin plug is provided and recommended for the instrument's safe operation. Use of any other adapter plug will decrease the performance of the instrument's safety features and possible injury (electrocution) to the operator may result.

- ★ Do not install the instrument in a location where is difficult to turn off the power and disconnect from the electrical receptacle.

- ★ The protection offered by the instrument's safety features may decrease or be nullified if the device is tampered with or not used as prescribed in this manual.

3. COMPONENT IDENTIFICATION

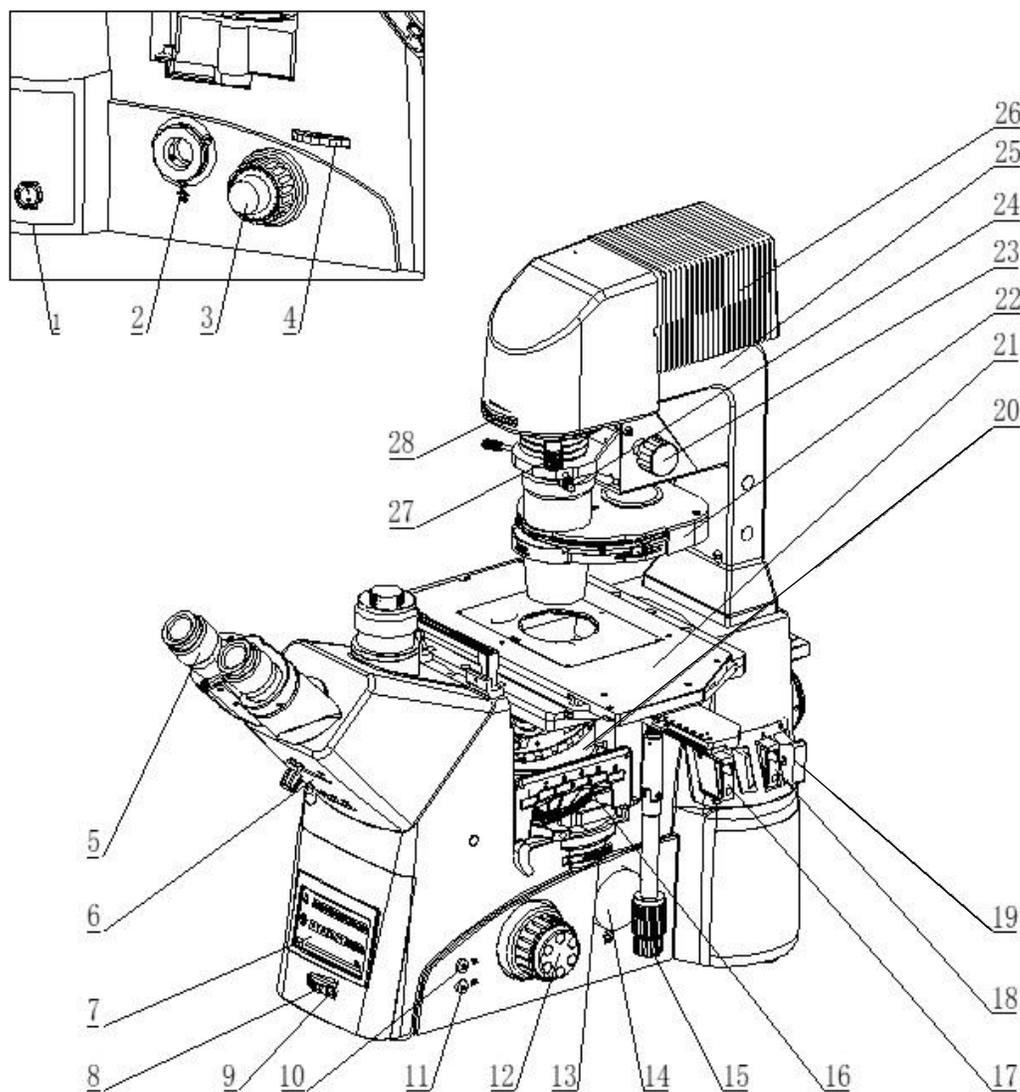
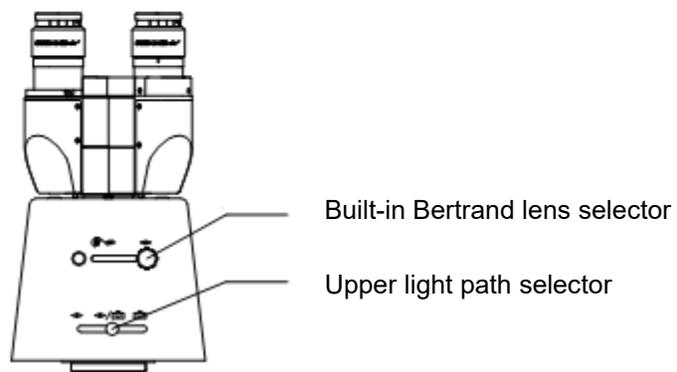
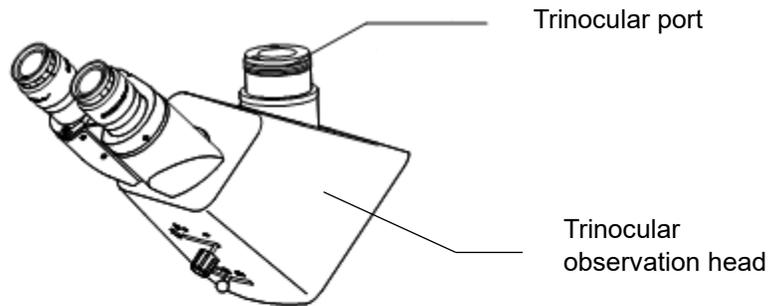


Fig. 1

- | | |
|---|---|
| 1. Power switch | 15. X-Y axis controls of sample stage |
| 2. Left camera port | 16. Reflected light/fluorescence filter cube turret |
| 3. Left coarse/fine focusing knob | 17. Reflected light field diaphragm |
| 4. Side port light path selector (left, right, binocular) | 18. Reflected light aperture diaphragm |
| 5. Eyepiece | 19. 3-hole filter slider |
| 6. Trinocular tube | 20. Objective nosepiece |
| 7. LCD Touch display screen | 21. Sample stage |
| 8. Brightness adjustment knob | 22. Condenser turret |
| 9. Caution light | 23. Condenser focusing knob |
| 10. Transmitted illumination shutter (On/Off) | 24. Condenser centering screw |
| 11. Reflected illumination shutter (Open/Closed) | 25. Transmitted illumination pillar |
| 12. Right coarse/fine focusing knob | 26. LED lamp house |
| 13. Magnification changer turret | 27. Filter holders |
| 14. Right camera port | 28. Transmitted light field diaphragm |

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4. APPLICATIONS

The EXI-600 inverted research microscope is primarily used to study living cells growing in tissue-culture dishes. The microscope can be configured with a variety of contrast techniques to enable observation, imaging and analysis such as brightfield (standard), phase contrast, differential interference contrast (“DIC”), and fluorescence, and is an ideal platform for integration with 3rd party accessories such as LED-based fluorescence illuminators, digital imaging cameras, motorized stages, motorized filter wheels, motorized focus, and manipulators. Please contact an ACCU-SCOPE salesperson to discuss your application and what peripherals are available to meet your needs.

5. ASSEMBLY

The EXI-600 will arrive partially assembled. Some parts and components may need to be installed and aligned before use and for optimal performance.

5.1. PREPARATION BEFORE ASSEMBLY

Tear down the packaging of the microscope frame and the accessories. Packaging includes the main frame, eyepieces, objectives, condenser, LED lamp house, stage, and other accessories such as filters, DIC blocks, dust-proof shield, tools (optional) and user manual. Optional and 3rd party accessories may be shipped in packages separate from the main packaging.



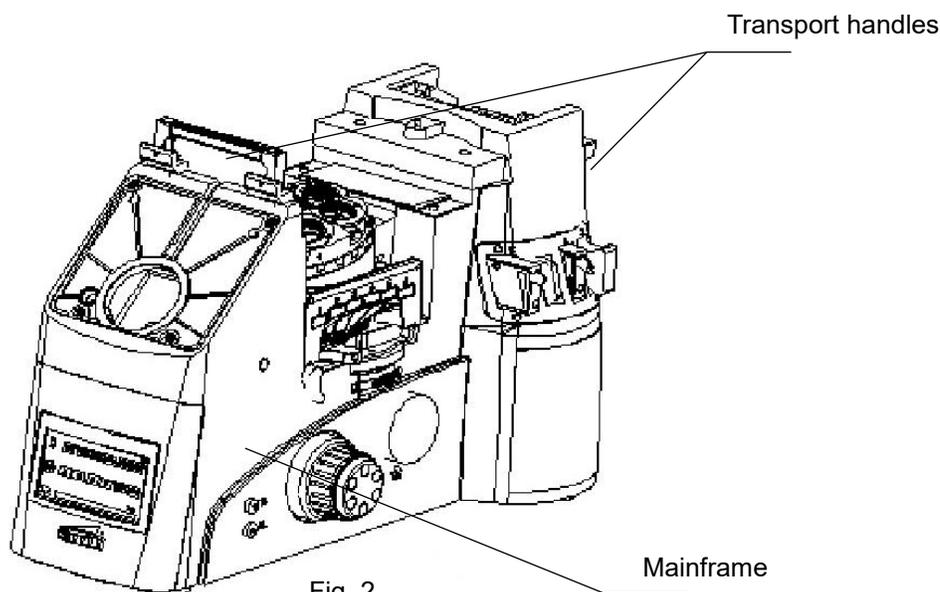
1. Open all the packaging, check the goods and confirm the accordance with the products you purchased.

★ Keep all dust caps, covers, tools, and user manuals in safe place.

2. Remove the transport handles shown in Fig. 2.

Put the mainframe on a sturdy, vibration-isolated platform and then remove each transport handle using a 4mm hexagon screwdriver.

★ Keep the transport handles and bolts should the microscope need to be moved at another time.



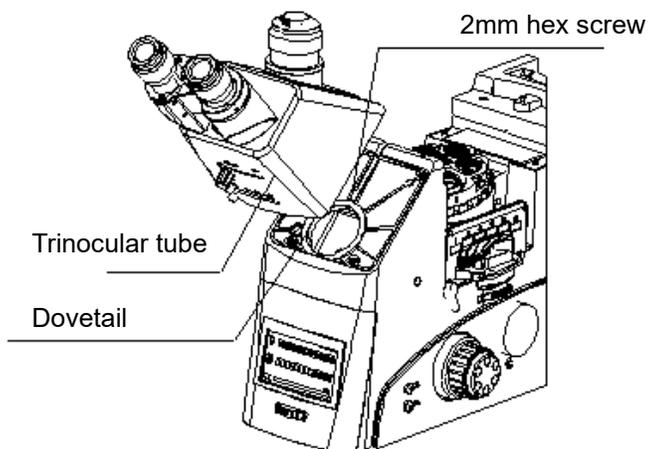
5.2. ASSEMBLY

5.2.1. Trinocular observation tube

Use a 2mm hex screwdriver to loosen the 2mm hex screw and remove the dust cap (do not discard; keep in safe place).

Align and place the dovetail of the trinocular tube into the receptacle on the frame. Tighten the 2mm hex screw to secure the trinocular tube onto the frame.

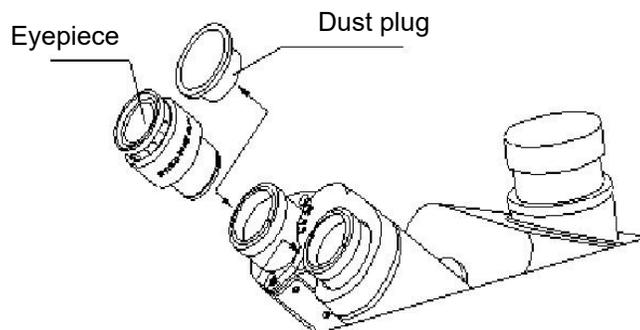
- ★ Hold the trinocular tube securely to avoid it falling during installation.
- ★ A slot is built into the mainframe through which the 2mm hex screwdriver will fit to tighten the 2mm hex screw.



5.2.2. Eyepieces

Remove the dust plugs from the eyetubes.

Insert the eyepieces into the eyetubes.



5.2.3. Transmitted illumination pillar

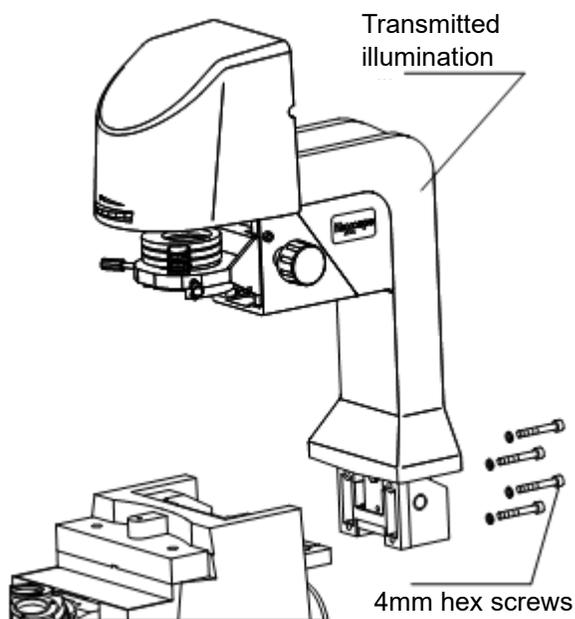
Use a 4mm hex screwdriver and remove the 4 hex screws from the back of the mainframe. These screws will be used again momentarily.

Hold the pillar above the mainframe with the base of the pillar directly above the mounting position.

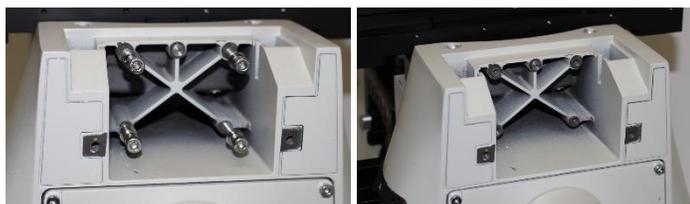
Align the guide pins with the slots in the frame.

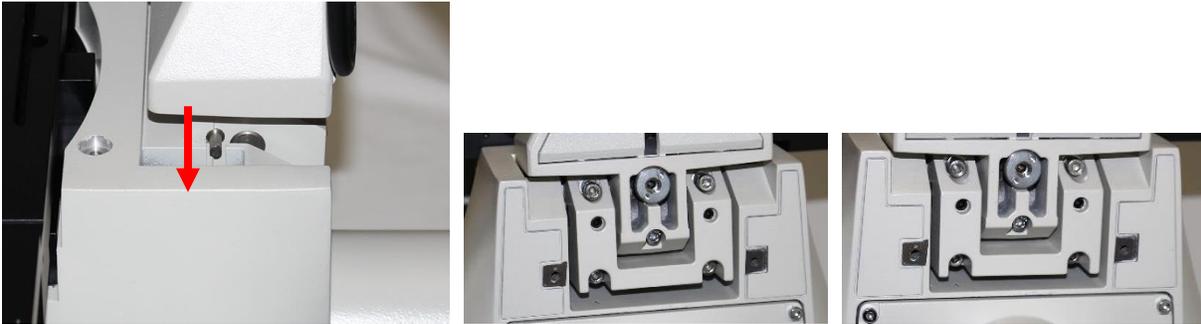
Slowly slide the pillar down into the slots. The pillar is not yet secured, but will not fall over.

Secure the pillar onto the frame using the same 4 hex screws from above.



★ *When installed properly, the pillar can be easily tilted backward and forward with one hand.*





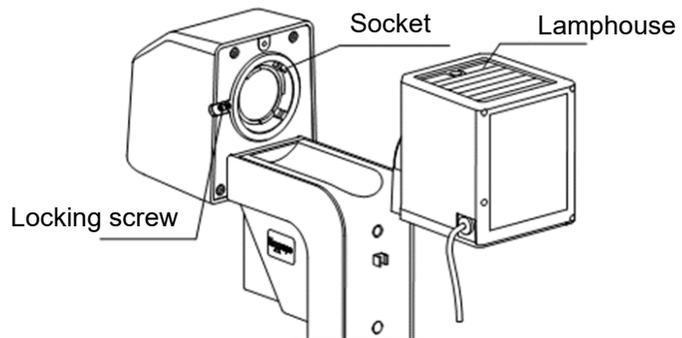
5.2.4. Transmitted LED Lamphouse

NOTE: The EXI-600 microscope typically ships with the transmitted LED lamphouse already installed. The instructions below may be useful for maintenance or replacement.

Insert the dovetail of the lamp house into the socket and ensure the dovetail is fully inserted. The lamphouse should be parallel to the bench.

Use a 2mm hex screwdriver and tighten the locking screw to secure the lamphouse to the socket.

Clip the power cord into the plastic clips on the back of the pillar.



5.2.5. Stage

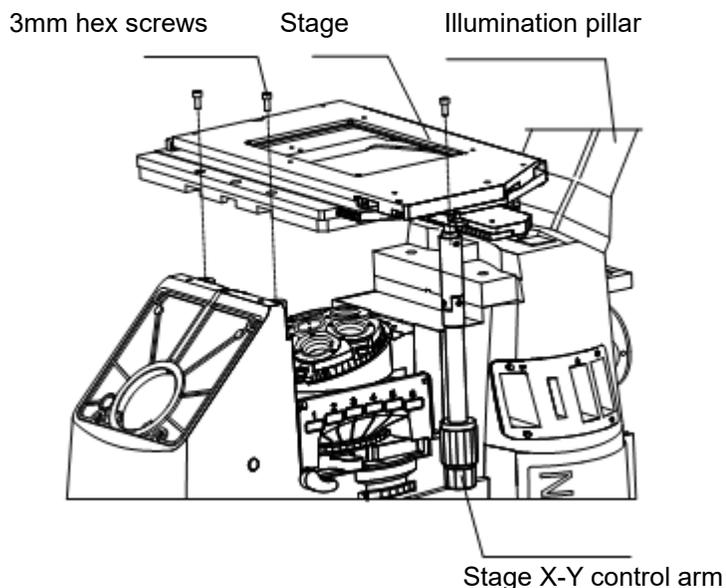
Tilt the transmitted illumination pillar into its back position.

Align the holes in the stage with the screw holes in the mainframe.

NOTE: The stage X-Y control arm should be positioned to the right.

NOTE: 2 screw holes are located in the front of the stage closest to the observation tube, and 1 screw hole centered in the back by the light pillar.

Use a 3mm hex screwdriver to secure the stage with the supplied 3mm hex screws.



5.2.6. Objectives

★ *Install the objectives from low magnification to high, beginning with the lowest position number on the nosepiece.*

★ *If installing different types of objectives (e.g., brightfield and phase contrast), install all of one type before installing the next.*

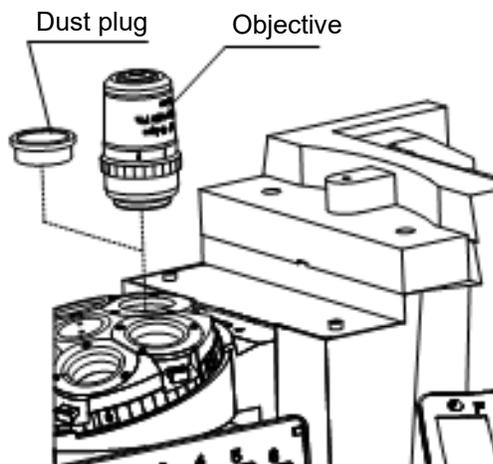
Remove a dust plug from the nosepiece. Screw an objective into the threaded opening.

★ *Note the position number on the nosepiece that corresponds to the objective just installed. This will be needed when programming the LCD screen later.*

★ *Do not over tighten the objective. Hand tight is sufficient.*

Repeat until all objectives are installed.

★ *To minimize dust from getting on the optics below, leave dust plugs in place where objectives are not installed.*



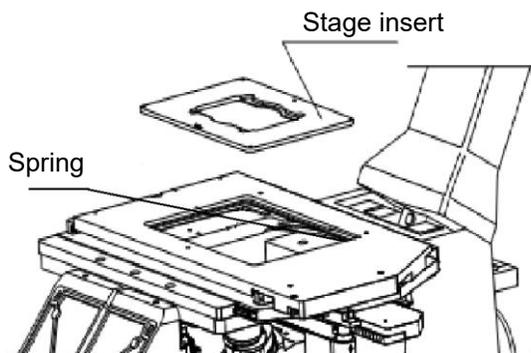
5.2.7. Stage inserts

- ★ *Stage inserts adapt the stage to accommodate various specimen holders such as microscope slides and multiwell plates.*

Align the stage insert close to the spring on the edge of the stage.

Press the stage insert into the stage opening until it is level and flush with the stage surface.

- ★ *Only one stage insert can be installed at a time, and this procedure allows for easy exchange of inserts based on need.*



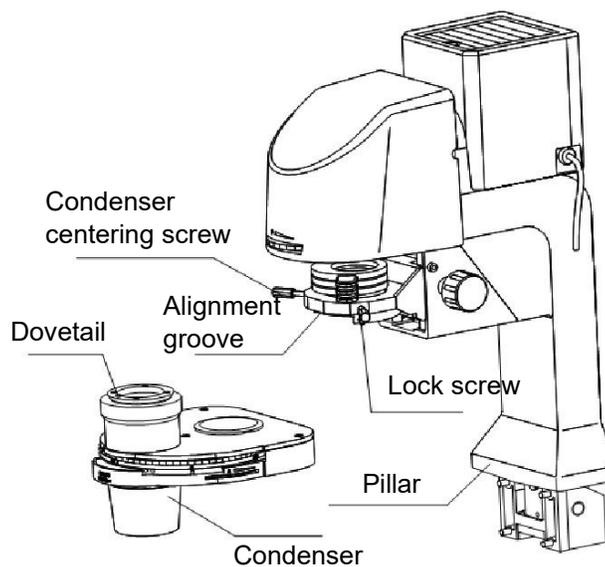
5.2.8. Turret condenser

- ★ *Install any additional inserts (e.g., DIC prism or phase annulus) prior to installing the turret condenser onto the transmitted light illumination pillar.*

Align the guiding pin on the top-front edge of the dovetail with the alignment groove on the front of the condenser carrier.

Slide the dovetail of the condenser into the condenser carrier. Ensure the dovetail is completely inserted into the carrier.

Secure the condenser by tightening the lock screw.



5.2.9. Differential Interference Contrast (DIC): Component installation

- ★ *DIC observation is highly sensitive to dust in the optical system. Use special care (e.g., wear dust-free nitrile or vinyl gloves) when handling DIC components to ensure they are free of dust and fingerprints.*
- ★ *Installation of DIC condenser prisms should be performed **ONLY** by ACCU-SCOPE-authorized service technicians. The instructions below are for informational purposes only.*

Insert or exchange a DIC prism in the condenser turret

Remove the condenser from the condenser carrier of the transmitted illumination pillar.

Remove the dust cover on the top of the condenser.

Rotate the condenser turret until either the DIC or DIC II position is centered in the opening.

Remove the retaining ring if present.

Set the DIC condenser prism into the opening. Be sure to align the groove of the prism with the pin in the opening, and that any writing is oriented upright ("40X", not "X04"). Ensure that the prism is completely seated in the opening.

DIC position: for 10X objective.

DIC II position: for 20X/40X/60X objectives.

Thread the retaining ring into place, and replace the dust cover.

Repeat as necessary for another DIC condenser prism.

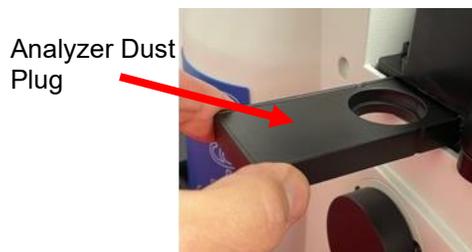
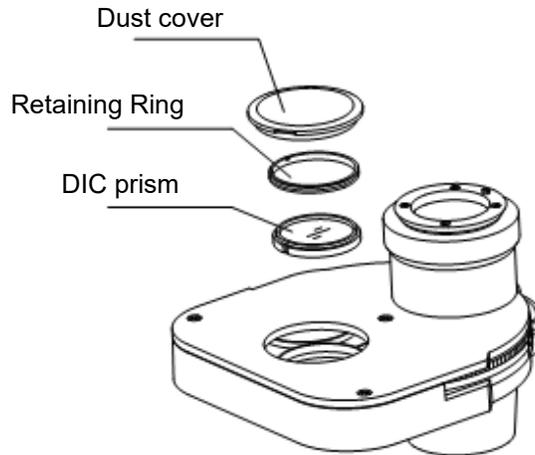
Mount the condenser back in the condenser carrier on the pillar.

Installing the analyzer and objective prisms

Remove the analyzer dust plug located under the fluorescence cube turret.

Insert the DIC analyzer slider.

- ★ *The analyzer slider has 2 positions (in/engaged and out/disengaged) indicated by*



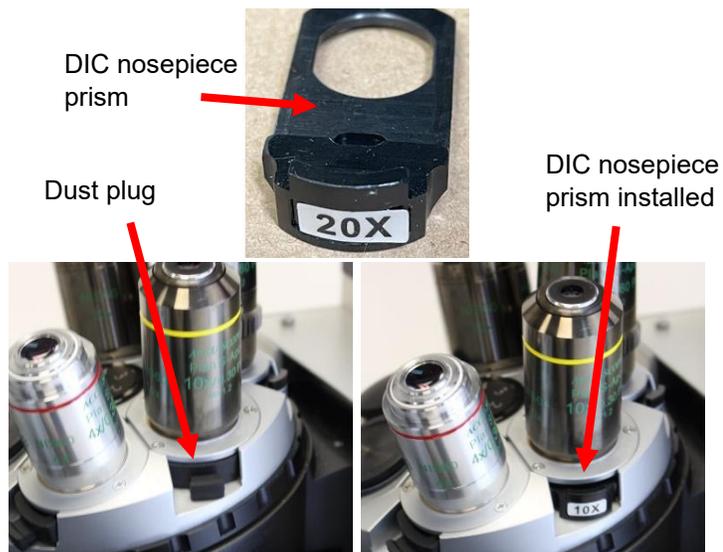
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detents. When engaged, the analyzer is aligned with the optical path. When disengaged, the analyzer is completely removed from the optical path.



Remove the rubber dust plug located just below the objective to be used for DIC observation.

Insert the DIC nosepiece prism that corresponds to the objective magnification.



5.2.10. Fluorescence filter cube turret (optional)

Install the filter cubes into the cube turret.

Insert the bottom of the “wings” of a cube into the metal clip.

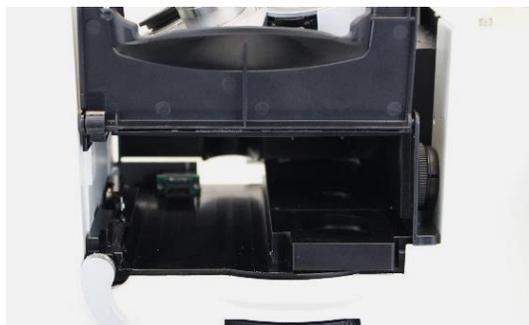
Gently push onto the back of the cube until the top of the wings click into the upper metal clips.



★ *NOTE: When installed correctly, there is no filter in the upper face of the filter cube (filters occupy the front and bottom positions, only, in addition to the beamsplitter).*

To insert the cube turret into the frame, turn the locking lever down.

Raise the cover to reveal the location into which the turret will be inserted.



Position the turret with the “handle” facing away from the right side of the microscope, and slowly slide the turret onto the track and into the opening. Ensure that the turret slides completely into the microscope.



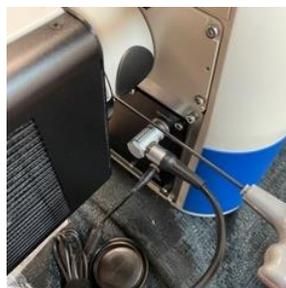
Close the cover and return the locking lever into the horizontal (locked) position.



5.2.11. Fluorescence light source (optional)

Align and insert the fluorescence light source adapter dovetail into the receiver on the back of the microscope (requires epi-fluorescence vertical illumination is already installed).

Secure the light source adapter by tightening the 2mm hex screw using a 2mm hex screwdriver.



5.2.12. UV shield (optional; only needed for fluorescence)

★ *NOTE: A UV shield is recommended for fluorescence microscopy.*

Align the holes in the UV shield brackets with the tapped holes in the top of the mainframe and closest to the observation tube. Using a 4mm hex screwdriver, insert and tighten the provided hex screws to secure the shield in place.



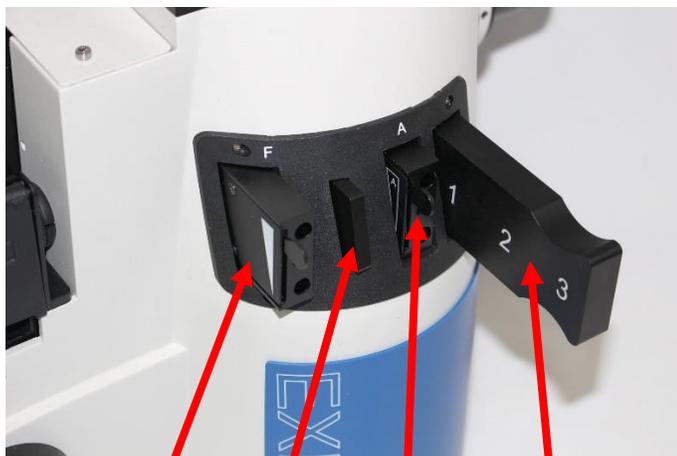
5.2.13. Fluorescence sliders (optional; fluorescence only)

Fully insert the Reflected light field aperture slider into the “F” slot of the mainframe. Note the orientation of the white triangle indicator for light throughput. Detents in the slider help identify the “out” and “in” positions.

Fully insert the Reflected light aperture diaphragm slider (indicated with an “A”) into the “A” slot of the mainframe. Detents in the slider help identify the “out” and “in” positions.

Insert the 3-position Neutral Density (“ND”) filter slider into the rear-most slot (to the right of the Aperture iris slider).

Starting adjustment for fluorescence: Set the Field and Aperture sliders to the top (fully open) positions. Using the detents on the ND filter slider, push in or pull out the slider until the middle (empty) position is in the light path – this position has no attenuation of fluorescence signal and is a good starting point for fluorescence observation.



Reflected light field diaphragm slider

Blank slider

Reflected light aperture diaphragm slider

Neutral density filter slider

5.2.14. Power supply

Connect the cable from the LED lamp house into the 3-pin power coupler labeled “OUTPUT” located on the back of the microscope. The lamp house power connector must be oriented in the 3 o'clock position. Press in gently to secure the connection.

To remove, simply grasp the power connector and pull straight back from the microscope.

Connect the power adapter to the power port labeled “INPUT” on the back of the mainframe.

Attach the slotted end of the power cord to the power adapter, and the plug to a suitable electrical receptacle.



5.2.15. Camera adapter

Remove dust cap from a camera port by pulling straight off.

Thread a camera adapter onto the camera.

- ★ *To avoid getting dust onto the camera sensor or inside the camera adapter, hold them both in a horizontal orientation when threading them together.*

Thread the camera adapter with attached camera onto the side port.

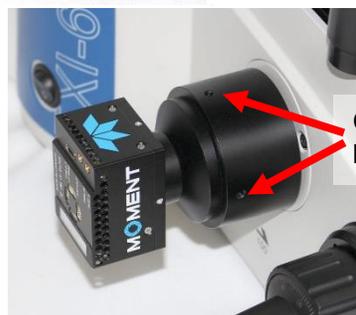
Orient Camera Image

- ★ *After installing the camera, the image to the monitor may be in a different orientation than seen through the eyepieces.*

Loosen the camera adapter locking screws (3 total) with a 2mm hex screwdriver.

Compare the image on the monitor with the image seen through the eyepieces, and rotate the camera until the orientation is the same.

Tighten the locking screws to secure the camera orientation.



Camera adapter locking screws

6. OPERATION

6.1. COMPONENT ADJUSTMENTS

6.1.1. Trinocular observation tube adjustments

Eyepiece diopter adjustment

Set the diopter adjustment on each eyepiece by rotating the eyepiece while holding the barrel. Align the "0" mark with the white dot.

Focus on a specimen on the stage until the image in one eyepiece is perfect.

Adjust the diopter of the other eyepiece until the image through that second eyepiece is good.

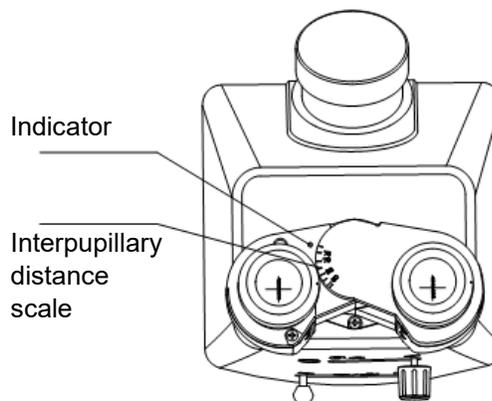
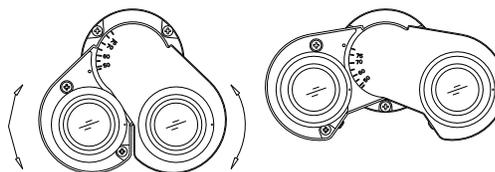
★ *The eyepieces can be adjusted by a maximum of ± 5 diopters.*

Adjust interpupillary distance

While viewing a sample, grasp and adjust the eyetubes up or down at the binocular hinge to increase or decrease the interpupillary distance (distance between the pupils of the observer's eyes).

Proper interpupillary distance results in a single uniform view of the specimen.

Interpupillary distance range:
55~75mm

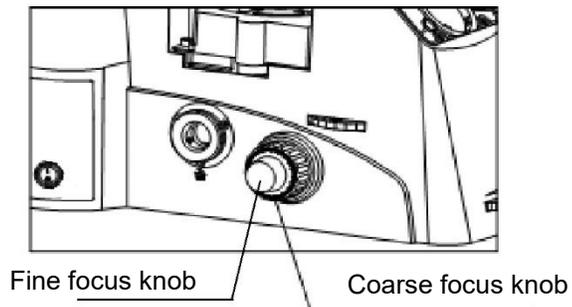


6.1.2. Coarse/Fine focus

Total focus movement range:
10mm

Course focus adjustment:
2mm/revolution

Fine focus adjustment:
0.2mm/rotation



- ★ *Rotation of the focus knobs away from the operator (i.e., left-side focus knobs move in counterclockwise direction) moves the objective nosepiece up. Rotation of the focus knobs toward the operator (i.e., left-side focus knobs move in clockwise direction) lowers the nosepiece.*
- ★ *Focusing knobs are located on both sides of the microscope. The left side displays a scale, 0.10mm/division.*

6.1.3. Condenser turret: Components

6-position condenser turret

B : Brightfield

Ph1 : Phase contrast for 10X/20X phase contrast objectives

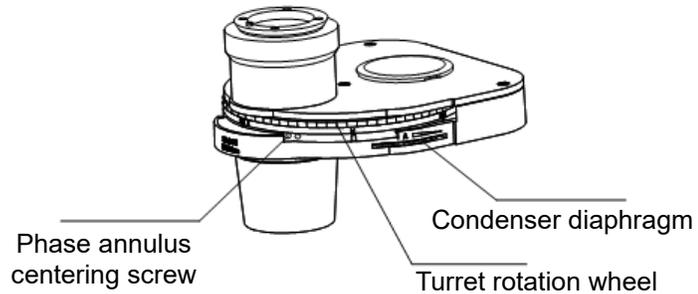
Ph2 : Phase contrast for 40X phase contrast objective

Ph3/L : Phase contrast position for 60X (Ph3) or 4X (PhL) phase contrast objective (only supports one phase annulus)

DIC : Differential Interference Contrast for 10X objective

DIC II : Differential interference Contrast for 20X/40X/60X objectives.

Adjust the aperture diaphragm by rotating the condenser diaphragm wheel.



- ★ The condenser aperture diaphragm is adjustable **ONLY** when the condenser turret is in the B, DIC and DIC II positions. **The aperture is automatically fully open ("auto-iris" feature) for Ph1, Ph2 and Ph3/L positions and cannot be adjusted.**



Change to another position on the condenser turret by rotating the turret rotation wheel. Rotate until the desired position faces front (i.e., B for brightfield).

6.1.4. Kohler Illumination: Aligning the condenser in the optical path

Centering the condenser

Set the condenser turret for brightfield observation ("B" position) and using the 10X objective, focus on a specimen.

Close the field diaphragm completely.

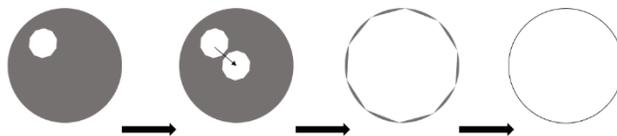
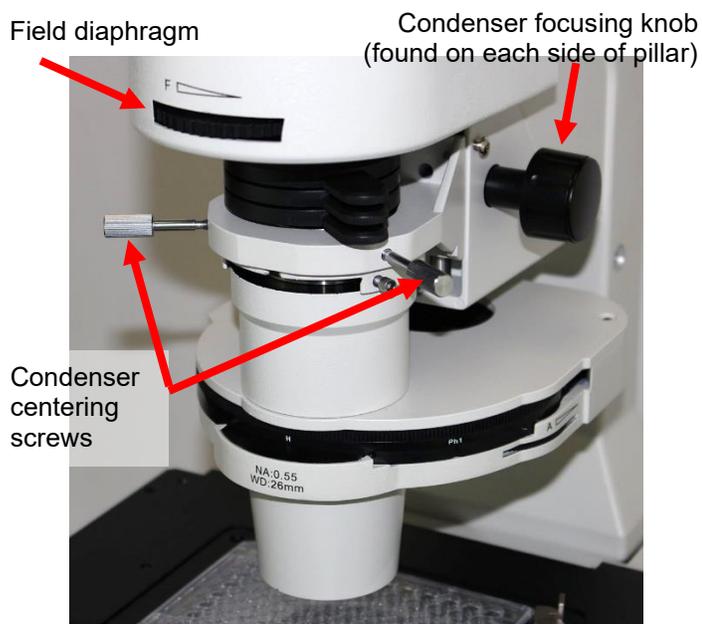
Using the condenser focusing knobs, adjust the condenser position up or down until the bright area seen in the specimen is seen sharply. Flat edges of the field diaphragm should become visible.

Using the condenser centering screws, center the bright area in the field of view by screwing in or out the centering screws.

Open the field diaphragm until the bright area fills most of the field of view.

Adjust condenser centration again using the centering screws.

Open the field diaphragm until it is outside the field of view.



Adjusting the condenser diaphragm

- ★ The condenser aperture diaphragm automatically adjusts (“auto-iris” feature) to be fully open in the Ph1, Ph2 and Ph3/L condenser turret positions. **DO NOT ADJUST THE CONDENSER APERTURE DIAPHRAGM WHEN IN Ph1, Ph2 or Ph3/L positions.**

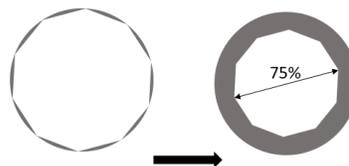
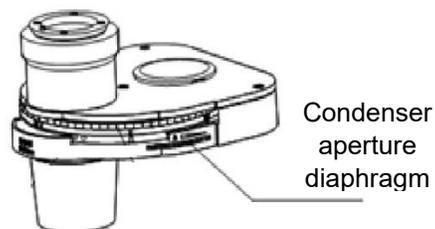
Turn the condenser turret to the brightfield “B” position.

Focus on a specimen.

Remove one eyepiece. If it is too bright, reduce illumination intensity so that the leaves of the condenser aperture diaphragm can be clearly distinguished.

Adjust the condenser diaphragm until the field of view is illuminated approximately 75% (open).

- ★ ***The condenser aperture position is directly related to the Numerical Aperture of the objective, and adjustment should be performed after every objective change.***



6.1.5. Condenser turret: Centering phase contrast annuli

- ★ *Phase contrast annuli ship preinstalled in the condenser turret and are pre-centered. Centering is recommended as some movement may have occurred during transit.*
- ★ *Phase contrast observation and centering of the phase contrast annuli requires phase contrast objectives. These can be readily identified by green markings on the objective barrel and include a marking that indicates the appropriate phase contrast annulus to use (i.e., Ph1 for 10X, Ph2 for 20X and 40X, Ph3/L (optional) for 60X or 4X, respectively).*

Place and focus on a specimen on the stage.

Change to the phase contrast objective for which the corresponding phase annulus will be centered.

Change the condenser turret to the phase annulus position (Ph1, Ph2, Ph3/L) that corresponds to the objective magnification.

The condenser diaphragm is fully open automatically.

Engage the built-in Bertrand lens by moving the black knob on the front of the observation tube to the "B" position.

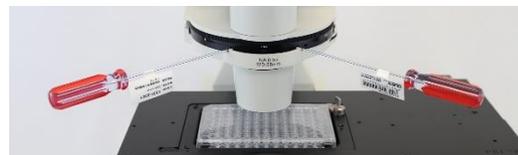
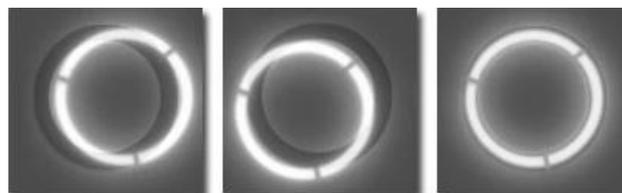
While looking through the eyepieces, two rings are visible and of nearly the same size – one dark and one bright.

- ★ *If the two rings are of vastly different sizes, the wrong phase contrast annulus is in position. Turn the condenser turret to the correct phase position.*

The black knob of the Bertrand lens slider may be turned to help focus the rings.

The dark ring is fixed and located in the objective – this ring cannot be moved.

Insert each of the two 1.5mm hex screwdrivers into the centering screws (left and right) of the condenser.



While looking through the eyepieces, slowly turn the centering screws to move the bright ring to be aligned with the dark ring.

Return the black knob on the observation tube to the normal viewing position .



6.1.6. Differential Interference Contrast (DIC): Component adjustment

Aligning and optimizing DIC components

- ★ *For optimal performance, the polarizer and analyzer must be properly aligned. The DIC prisms are fixed and cannot be adjusted.*
- ★ *The DIC condenser polarizers are combined with the DIC condenser prisms.*
- ★ *BEFORE proceeding with DIC setup, perform Köhler illumination per section 6.1.4 above.*

Rotate the condenser turret to the DIC or DIC II position.

Select an objective with DIC prism that corresponds to the DIC condenser prism selected above (i.e., 10X with DIC, 20X or 40X with DIC II).

Temporarily, remove the DIC nosepiece prism.

Focus on a specimen.

Engage/insert the analyzer fully.

Loosen slightly the thumbscrew on the analyzer gauge.

While looking through the eyepieces, rotate the gauge on the analyzer (wheel) until the image is black or as “black” as possible. This position indicates “maximum extinction” when the polarizer (in the DIC condenser prism) and the analyzer are oriented at 90° to each other.

Gently tighten the thumb screw on the analyzer gauge to hold the extinction position.

Replace the DIC nosepiece prism.

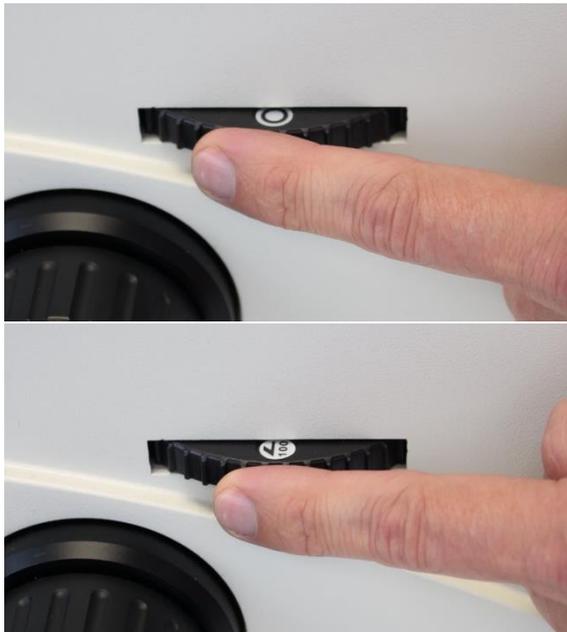


6.1.7. Side port light path selector

The side port light path selector allows the specimen image to be observed from the eyepieces, or the left or right camera ports.

The side port selector wheel has 3 positions with different light splitting conditions:

-  100% eyes : 0% camera port
-  20% eyes : 80% right-side camera port
-  0% eyes : 100% left-side camera port



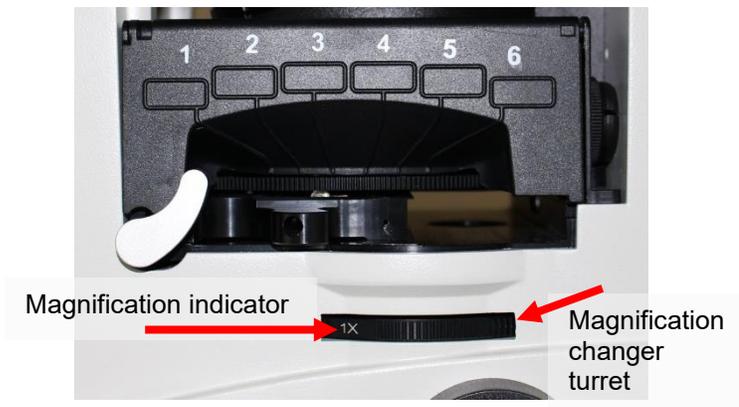
6.1.8. Magnification changer

Rotate the magnification changer to change the magnification for observation. Available magnifications:

1X : Does not alter the magnification of the view

1.5X : Increases the magnification by 1.5X

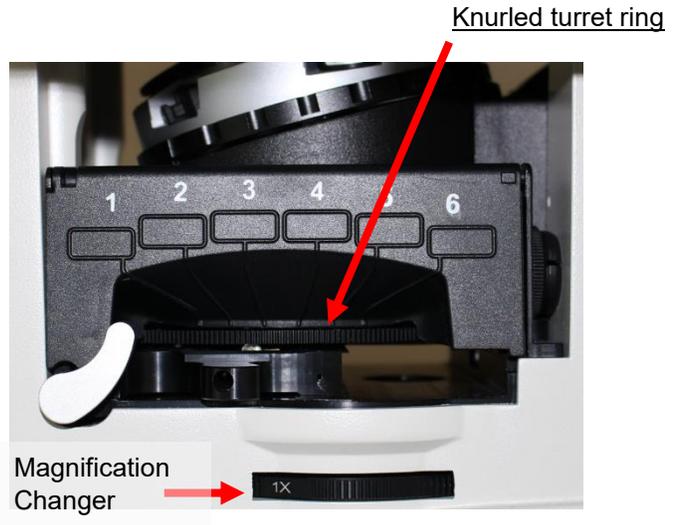
Blank (no label) : This position is reserved for future feature development.



6.1.9. Fluorescence components

Fluorescence Filter Cube Turret.

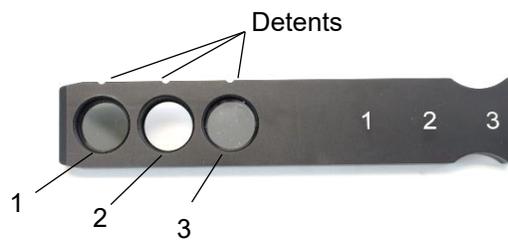
Rotate turret using the knurled ring. A positive detent indicates that the turret is in a filter cube position. The LCD screen displays the position of the turret and filter cube that is in the light path for fluorescence observation.



Neutral Density "ND" filter slider.

Insert slider to desired position to attenuate fluorescence illumination intensity.

- Position 1: 50% Transmission
- Position 2: 100% Transmission
- Position 3: 25% Transmission

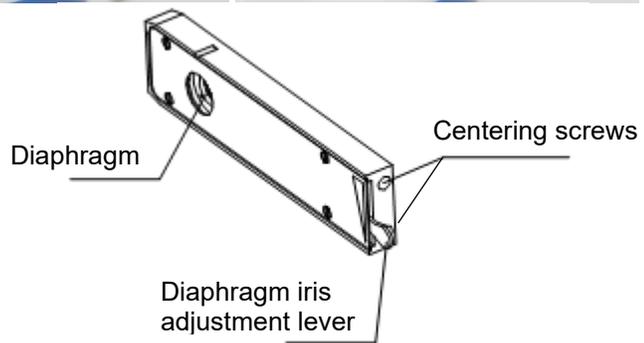


Field diaphragm: Controls the size of the illuminated field.

Adjust the diaphragm opening by moving the diaphragm iris adjustment lever up to open, or down to close.

The field diaphragm iris can be centered in the light path by adjusting the two centering screws using a 3mm hex screwdriver.

1. Place a specimen or piece of paper on the stage.
2. While viewing the sample under fluorescence illumination, close the field diaphragm part way



Iris open:



Iris closed:

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- until the leaves of the iris can be seen in the field of view.
3. Adjust the centering screws until the iris is centered in the field.
 4. Open the diaphragm iris completely.

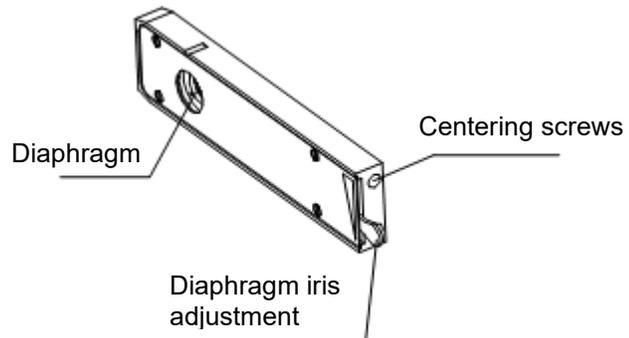
Aperture diaphragm: Regulates the intensity of illumination without affecting the size of the field.

Adjust the diaphragm opening by move the diaphragm iris adjustment lever up to open, or down to close.

- ★ *The aperture diaphragm is pre-centered at the factory and further adjustment should not be necessary.*

Blank slider.

Ensure that the blank slider is fully inserted into the slot. The detent will engage.



6.1.10. Programming the LCD panel

After programming, the LCD panel displays the current status of objective position, fluorescence filter cube turret position, and transmitted light illumination level.

Objective positions: Each of the six positions on the intelligent nosepiece can be programmed into the LCD panel. When in program mode, choose the magnification that corresponds to the objective in that position on the nosepiece.

Note that the intelligent nosepiece does not automatically recognize the presence of an objective.

NOP = No Object Present

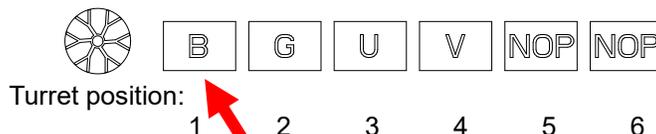
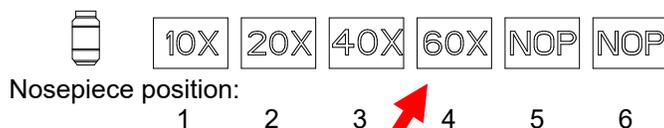
Fluorescence filter cube turret positions: Each of the six positions on the intelligent filter cube turret can be programmed into the LCD panel. When in program mode, choose the magnification that corresponds to the filter cube in that position of the turret.

Arrows on the turret indicate the position as recognized by the LCD panel. Make a note of these when the filter cubes are inserted into the turret.

RECOMMENDATION: Choose an LCD panel button indicator that corresponds to the excitation color of the filter cube being programmed (e.g., “B” for blue, “G” for green, “U” for UV or DAPI, “V” for visible or brightfield, etc.).

Note that the intelligent turret does not automatically recognize the presence of a filter cube.

NOP = No Object Present



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This bar indicates the percent of maximal intensity of transmitted light.

Adjust intensity using the brightness adjustment wheel located below the LCD screen.

Transmitted light intensity



Program the objective positions and fluorescence filter cube positions using the Settings “gear” icon .

Press the Settings icon. It will change to a bubble appearance.

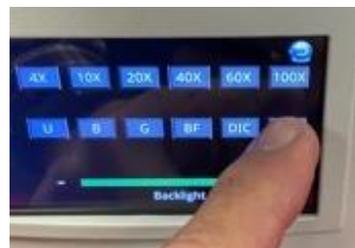
Press a button for objective or fluorescence turret position to program.

Choose from the available options.

Press another button to program it.

When finished, press the bubble icon to accept the changes and the bubble icon will change back to the Settings “gear” icon.

These settings are saved even if power to the microscope is turned off.



6.2. MICROSCOPE OPERATION

6.2.1. Transmitted Light Methods

6.2.1.1. Brightfield observation – Instrument configuration

Component	Position
Transmitted illumination shutter	On (open) , light is coming out of the bottom of the condenser
Upper light path selector	
Lower light path selector	
Condenser turret position	“B”
Magnification changer	1X
Fluorescence cube turret (if installed)	Choose an open position (no filter cube installed)

1. Place a specimen on the stage.
2. Focus on the specimen using the focus knobs.
3. If desired, change side port light path selector or trinocular tube upper light path selector to a camera port for imaging.

6.2.1.2. Phase contrast observation – Instrument configuration

Component	Position
Transmitted illumination shutter	On (open), light is coming out of the bottom of the condenser
Upper light path selector	
Lower light path selector	
Condenser turret position	“Ph1” / “Ph2” / “Ph3/L”
Magnification changer	1X
Fluorescence cube turret (if installed)	Choose an open position (no filter cube installed)

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Objective(s)	<p>Use phase contrast objectives, identifiable by the green writing on the objective barrel.</p> <p>Match the objective with the condenser turret position:</p> <p style="padding-left: 40px;">Ph1 : 10X/20X objectives</p> <p style="padding-left: 40px;">Ph2 : 40X objective</p> <p style="padding-left: 40px;">Ph3/L : 60X or 4X objective, respectively</p>
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1. Place a specimen on the stage.
2. Focus on the specimen using the focus knobs.
3. If necessary, recenter the phase contrast annulus in the condenser turret. See section 6.1.5.
4. If desired, change side port light path selector or trinocular tube upper light path selector to a camera port for imaging.

6.2.1.3. Differential interference contrast (“DIC”) observation – Instrument configuration

Component	Position
Transmitted illumination shutter	On (open), light is coming out of the bottom of the condenser
Upper light path selector	
Lower light path selector	
Condenser turret position	“DIC” / “DIC II”
Magnification changer	1X
Fluorescence cube turret (if installed)	Choose an open position (no filter cube installed)
Objective(s)	<p>Match the objective with the condenser turret position:</p> <p style="padding-left: 40px;">DIC : 10X objective</p> <p style="padding-left: 40px;">DIC II : 20X/40X/60X objectives</p>
Objective DIC prism	Ensure prisms are installed in slots below the corresponding objectives
Rotatable analyzer	Fully inserted position

1. Place a specimen on the stage.
2. Focus on the specimen using the focus knobs.
3. If necessary, reconfirm extinction of the condenser DIC component and the rotatable analyzer. See section 6.1.6.
4. If desired, change side port light path selector or trinocular tube upper light path selector to a camera port for imaging.

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6.2.2. Reflected Light Methods

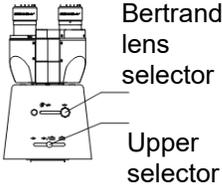
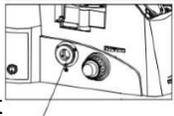
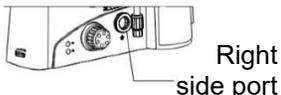
6.2.2.1. Fluorescence observation – Instrument configuration

Component	Position
Transmitted illumination shutter	Off (closed), no light is coming out of the bottom of the condenser
Reflected light shutter	On (open)
Reflected light field diaphragm	Fully open
Reflected light aperture diaphragm	Fully open
Neutral density filter slider	Open position
Fluorescence light source	On and shutter open
Upper light path selector	
Lower light path selector	
Condenser turret position	Any position. Condenser is not being used during fluorescence observation. It is recommend to tilt the light pillar to the back position to avoid reflection and autofluorescence of the transmitted LED light source.
Magnification changer	1X
Fluorescence cube turret (if installed)	Choose an appropriate filter cube for the specimen
Objective(s)	Any, as provide appropriate magnification of the specimen

1. Confirm that fluorescence excitation illumination is coming from the objective.
2. Place a specimen on the stage.
3. Focus on the specimen using the focus knobs.
4. Change fluorescence cube turret position to a different filter cube and observe again.
5. If desired, change side port light path selector or trinocular tube upper light path selector to a camera port for imaging.

6.3. MICROSCOPY IMAGING

Use the light path selector to send image information to the desired camera port. Camera port options include:

Camera Imaging Port	Microscope Location	Light Path Selectors	Selector Position
Trinocular port	 <p>Trinocular port</p>	 <p>Bertrand lens selector Upper selector</p>	<p>Bertrand lens selector: </p> <p>Upper selector:  = 50% eyes : 50% camera port  = 0% eyes : 100% camera port</p> <hr/> <p>Bertrand lens selector: </p> <p>Upper selector: No observation options available.</p>
Left side port	 <p>Left side port</p>		 = 0% eyes : 100% camera port
Right side port	 <p>Right side port</p>		 = 20% eyes : 80% camera port

1. Focus on specimen.
2. Change light path selector to desired camera port. Image should now appear on computer monitor or camera display.
3. Adjust camera settings per camera manufacturer or software developer's recommendations.
4. Observe and/or acquire images as desired.

7. TECHNICAL SPECIFICATION

7.1. MAIN TECHNICAL SPECIFICATION

Optical system	Infinity corrected system
Observation tube	Binocular tube, inclined 45°, adjustable interpupillary distance
Eyepiece	WF10X/22mm
Objective nosepiece	Sextuple nosepiece with DIC prism slots
Objectives	Brightfield/DIC/Fluorescence: 4X/10X/20X/40X/60X Plan semi-Apochromat; 10X/20X/40X/60X oil/100X oil Plan Apochromat Phase contrast: 4X/10X/20X/40X/60X Plan semi-Apochromat Modulation contrast: 10X/20X/40X S-Plan Fluor
Focusing	Coaxial coarse and fine focus Coarse focusing: 2mm/rotation Fine focusing: 0.2mm/rotation Stroke (from focal point at stage):7mm up, 2mm down
Stage	Range of movement: 135mm (X direction), 80mm (Y direction)
Illumination	10W LED, pre-centered, continuous variable intensity
Condenser	Six position, rotatable, focusable, NA 0.55, WD 26mm
Operating environment	<ul style="list-style-type: none"> • For indoor use only • Max altitude: 2000m • Environment temperature: 5°C~40°C (41°F~109°F) • Max relative humidity: 80% @ 31°C (88°F) Linear decrease corresponding to temperature 70% @ 34°C (93°F) 60% @ 37°C (99°F) 50% @ 40°C (104°F) • Pollution level: 2 • Power supply: ~220V ± 10%, 50/60HZ • Consumed power: 100W • Fuse: T5A/250V, Ø 5x20mm • Air pressure: 80kPa~106kPa • Overvoltage category: II

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7.2. OBJECTIVE SPECIFICATIONS

Class*	Magnification	Numerical Aperture (NA)	Correction Collar	Working Distance (mm)	Glass Slide Thickness (mm)
BF/DIC/FL: Plan APO	10X	0.45		4.0	0.17
	20X	0.75		1.1	0.17
	40X	0.95	0.11-0.23	0.21	0.17
	60X oil	1.40		0.14	0.17
	100X oil	1.45		0.13	0.17
BF/DIC/FL: Plan semi-APO	4X	0.13		17.0	1.2
	10X	0.30		7.4	1.2
	20X	0.45		8.0	1.2
	40X	0.60		3.3	1.2
	60X	0.70		1.9	1.2
PH/BF/DIC: Plan semi-APO	4X	0.13		17.0	1.2
	10X	0.30		7.4	1.2
	20X	0.45	0-2	7.5-8.8	1.2
	40X	0.60	0-2	3.0-4.4	1.2
	60X	0.70	0.1-1.3	1.8-2.6	1.2
MC/BF: sPlan-FL	10X	0.25		10.5	1.2
	20X	0.45	0-2	8.1-7.0	
	40X	0.60	0-2	3.6-2.8	

*BF = Brightfield

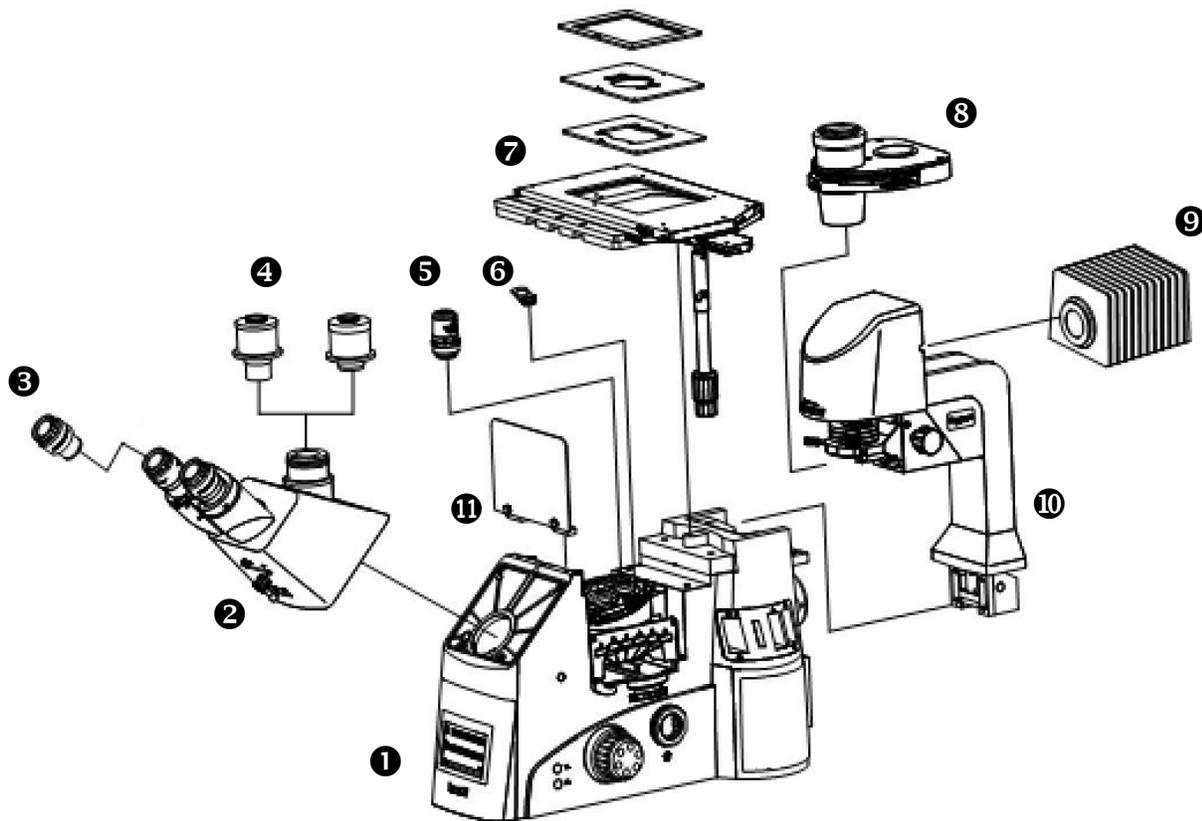
DIC = Differential interference contrast

FL = Fluorescence

PH = Phase contrast

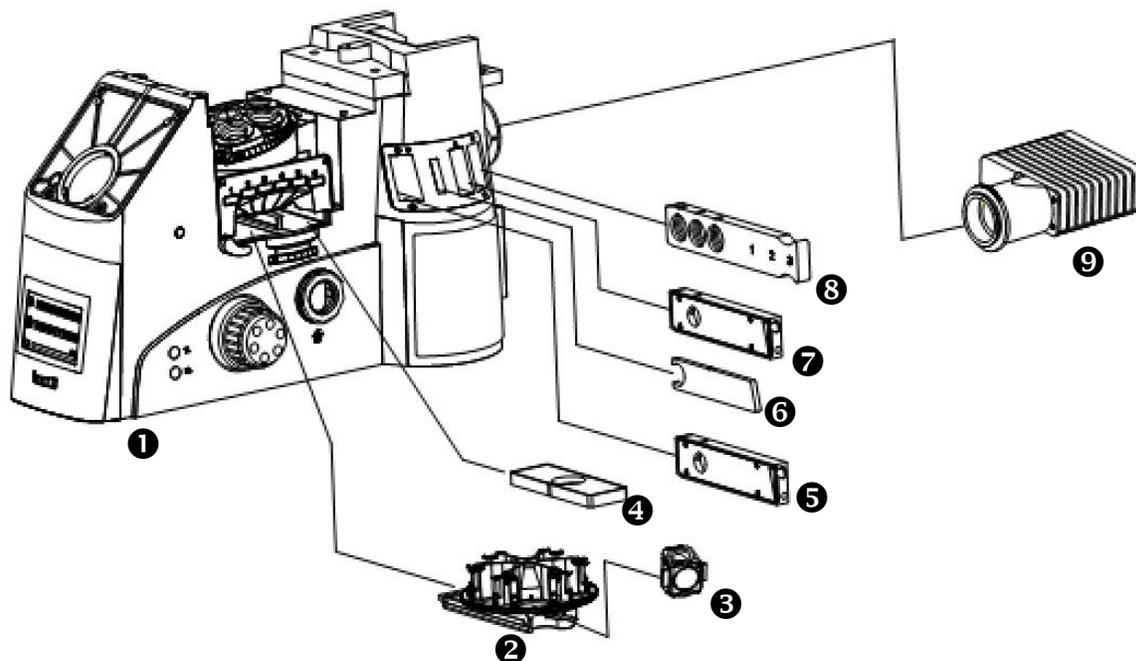
MC = Modulation contrast

8. EXI-600 CONFIGURATION DIAGRAM



- | | |
|--------------------------------|-------------------------------------|
| 1. Main microscope frame | 7. Stage |
| 2. Trinocular observation tube | 8. Condenser |
| 3. Eyepiece | 9. LED lamp house |
| 4. Camera adapter | 10. Transmitted illumination pillar |
| 5. Objective | 11. UV shield |
| 6. DIC nosepiece prism | |

9. EXI-600 FLUORESCENCE CONFIGURATION DIAGRAM



- | | |
|------------------------------------|----------------------------------|
| 1. Main microscope frame | 6. Blank slider |
| 2. Fluorescence filter cube turret | 7. Aperture diaphragm slider |
| 3. Fluorescence filter cube | 8. Neutral Density filter slider |
| 4. Analyzer slot blank slider | 9. Fluorescence light source |
| 5. Field diaphragm slider | |

10. TROUBLESHOOTING

Under certain conditions, performance of this unit may be adversely affected by factors other than defects. If a problem occurs, please review the following list and take remedial action as needed. If you cannot solve the problem after checking the entire list, please contact your local dealer for assistance.

10.1. OPTICAL

Problem	Cause	Corrective Measure
Illumination is turned on, but the field of view is still dark.	The LED bulb is burned out.	Replace with new 5W LED lamp.
	Illumination brightness is adjusted too low.	Turn up light intensity. Check LCD panel for brightness level.
	There are too many filters in the light path absorbing light.	Remove unnecessary filters from the light path (e.g., polarizers).
The field of view has a shadow or is unevenly illuminated.	The objective nosepiece is not fully in position.	Turn the nosepiece so that the detent is engaged in a position.
	Filter is not fully inserted or removed.	Push the filter all the way in, or remove it from the light path.
	Condenser turret position is not fully engaged.	Rotate the condenser turret until it sets completely into a position as felt by the detent.
Dirt or dust in the field of view.	Dirt or dust in the light path, e.g., on the eyepiece, condenser, objective, collector lens or specimen	Carefully clean the optics and specimen slide/vessel. Observe again and repeat until view is free of dirt.
A ghost image of the sample is observed.	Condenser aperture diaphragm is too closed.	Open the condenser aperture diaphragm.
Poor image quality. Low resolution Low contrast	Microscope optical and illumination components are not properly or optimally aligned.	Perform Köhler illumination.
	The objective nosepiece is not in the correct position.	Turn the nosepiece until the correct objective is in position.
	Condenser aperture diaphragm is too small or too big for brightfield observation.	Adjust the condenser aperture diaphragm according to Köhler illumination.
	Lenses or sample in the optical path (condenser, objective, eyepiece or culture dish) is dirty.	Carefully clean the optics and specimen slide/vessel.
	The thickness of culture dish bottom is over 1.2mm.	Use sample slides and vessels with thickness $\leq 1.2\text{mm}$.
	A brightfield objective is being used for phase contrast observation.	Change to a phase contrast objective.
	The phase contrast annulus (ring) does not match the	Ensure the correct phase contrast position on the

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	objective phase ring.	condenser turret is being used. Rings should be of same size when viewed in the eyetube (remove eyepiece first).
	The light ring is not centered on the dark ring during phase contrast.	Center the phase contrast annulus in the condenser turret using the centering screws.
	The phase contrast light and dark rings deviate when observing the edge of a tissue culture vessel.	Move the sample vessel to restore the phase contrast effect.
Image is blurry on one side of the field of view.	The objective nosepiece is not completely in position.	Turn the nosepiece until the correct objective is in position.
	The sample slide or vessel is not correctly positioned on the stage.	Ensure that the stage is securely installed on the microscope frame. Ensure that any stage plate inserts are completely installed and flush with the stage surface. Ensure that the sample slide or vessel is completely flat on the stage.
	The optical performance (e.g. flatness of the culture dish bottom) is poor.	Use a culture vessel with good flatness. Confirm quality of the culture vessel before seeding cells.

10.2. ELECTRICAL

Problem	Cause	Corrective Measure
LED bulb does not light.	Power not connected.	Check that the power cable from the LED lamp house is connected to the microscope frame, the microscope is connected to the power adapter, the power adapter is connected to a receptacle, and the power switch on the microscope is in the on position.
	Transmitted light shutter in engaged.	Press the transmitted light shutter button (TL) to disengage the shutter.
	LED bulb failed.	Replace the LED bulb.
Brightness is insufficient.	Light intensity adjustment too low.	Increase the light intensity of the LED. Observe the light intensity status on the LCD panel.
	Filters or other components in the light path.	Remove any filters, sliders, polarizers, etc. that are not being

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		used for the observation technique from the light path.
Light flickers.	Poor electrical connection.	Check all power cables to the LED lamp house and microscope are firmly attached.

OBSERVATION TUBE

Problem	Cause	Corrective Measure
The view with one eye does not correlate with the view with the other eye.	Interpupillary distance between eyepieces is incorrect.	Adjust the interpupillary distance of the binocular eyetubes until a single, overlapping image is observed when viewing with both eyepieces simultaneously.
	Eyepiece diopter is not properly adjusted.	Adjust the diopter of one eyepiece so that the focus of the image matches that seen through the other eyepiece.
	Still adapting to viewing through the eyepieces of this microscope.	It takes time to adjust to a different microscope. Try looking around the periphery of the field of view before concentrating on the specimen. Looking far away from the microscope (e.g., at an object across the room) before and during microscope use can aid eyes in acclimating to a microscope (and reduce fatigue).

10.3. MICROSCOPY IMAGING

Problem	Cause	Corrective Measure
Image not in focus.		
Edge or periphery of image is blurry compared to rest.		
Ghost image of window, interior room light, etc. appears in camera image.	Light entering eyepieces and merging with specimen image.	Cover eyepieces when imaging.
		Move camera to different camera port that allows 100% of light to go to the camera.

11. MAINTENANCE

1. Use gauze to gently wipe glass components. If you need to remove fingerprints and oily residue, slightly dampen the gauze first with a mixture of alcohol and diethyl ether (3:7) or dimethylbenzene before wiping.
 - ★ Diethyl ether and alcohol are both extremely flammable. DO NOT get them near open flames or any electrical components such as power switches. Always use these chemicals in well ventilated rooms.
2. Do not use organic solvents to wipe any non-optical components. If you want to clean these components, use a soft, dust-free cloth, moisten with a neutral cleaner in water, and wipe.
3. If the microscope is wet by liquid during operation, immediately turn off the power supply before cleaning up the liquid.
4. Do not disassemble any microscope components. This will immediately void any warranty and the performance and function of the microscope may be affected.
5. If not filling every position on the nosepiece with an objective, ensure that dust covers are installed in the unoccupied positions to prevent dust from entering the microscope body and collecting on other optics and components.
6. Cover the microscope with a dust cover when not in use. Wait until the lamp house cool before covering the microscope with the dust cover.
7. Any repair or replacement of components of this product should be executed by our company or an authorized ACCU-SCOPE service provider.

12. LIMITED MICROSCOPE WARRANTY

This microscope and its electronic components are warranted to be free from defects in material and workmanship for a period of five years from the date of invoice to the original (end user) purchaser. LED lamps are warranted for a period of two years from the date of original invoice to the original (end user) purchaser. The mercury power supply is warranted for a period of one year from the date of invoice to the original (end user) purchaser. This warranty does not cover damage caused in-transit, misuse, neglect, abuse or damage resulting from improper servicing or modification by other than ACCU-SCOPE approved service personnel. This warranty does not cover any routine maintenance work or any other work, which is reasonably expected to be performed by the purchaser. Normal wear is excluded from this warranty. No responsibility is assumed for unsatisfactory operating performance due to environmental conditions such as humidity, dust, corrosive chemicals, deposition of oil or other foreign matter, spillage or other conditions beyond the control of ACCU-SCOPE INC. This warranty expressly excludes any liability by ACCU-SCOPE INC. for consequential loss or damage on any grounds, such as (but not limited to) the non-availability to the End User of the product(s) under warranty or the need to repair work processes. Should any defect in material, workmanship or electronic component occur under this warranty contact your ACCU-SCOPE distributor or ACCU-SCOPE at (631) 864-1000. This warranty is limited to the continental United States of America. All items returned for warranty repair must be sent freight prepaid and insured to ACCU-SCOPE INC., 73 Mall Drive, Commack, NY 11725 – USA. All warranty repairs will be returned freight prepaid to any destination within the continental United States of America, for all foreign warranty repairs return freight charges are the responsibility of the individual/company who returned the merchandise for repair.

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